



A Review on Biomedical Properties of The Antioxidant Luteolin

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Abstract: 3', 4', 5, 7-tetrahydroxyflavone, luteolin, which is present in Onions, Broccoli and other dietary supplements, is an important flavinoid, present in our daily nutrition in less than 1 mg/day. Various Epidemiological studies have led to the discovery of the anti-inflammatory and anti-carcinogenic effects of luteolin. These properties are partly due to its anti-oxidant and free radical scavenging capacities. Luteolin was found to delay or block the development of cancer cells *in vitro* and *in vivo* by protection from carcinogenic stimuli, by the inhibition of tumor cell proliferation, by the arrest of cell cycle and by the induction of apoptosis through the suppression of cellular pathways, which induce the tumor suppressor p53. Luteolin was found to be the most effective flavinoid in inhibiting the tumor cell proliferation. Luteolin is being used to cure skin cancer. In this review, we focus on the reasons for the anticancer role and the molecular mechanisms which favours this property.

Keywords: Luteolin, apoptosis, p53, skin cancer, Pharmacology.

Introduction

Luteolin, 3', 4', 5, 7-tetrahydroxyflavone, belongs to a group of compounds, that occur naturally, called flavonoids. Flavonoids are polyphenols that play an important role in defending plant cells against microorganisms, insects, and UV irradiation [1]. It has been found that flavonoids possess anti-cancer properties [2, 3]. There is good evidence suggesting that flavonoids contribute to the cancer-protective effect of fruits and vegetable food [4]. Flavonoids acts by blocking several activities in the progression of carcinogenesis, including cell transformation, invasion, metastasis and angiogenesis, through inhibiting specific kinases, reduction of transcription factors, by cell cycle regulation, and by inducing apoptosis [5].

The structure of luteolin consists of a C6-C3-C6 structure and has two benzene rings (A, B) a third, oxygen-containing (C) ring, and a 2-3 carbon double bond. Luteolin also possesses hydroxyl groups at carbons 5, 7, 3', and 4' positions as in (Fig.1) [6]. The hydroxyl moieties and 2-3 double bond are important structure features in luteolin that are associated with its biochemical and biological activities such as anti-inflammatory and anti-carcinogenic activities [7].

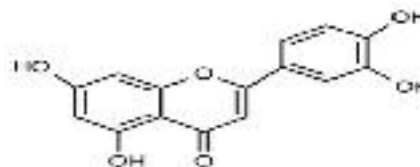


Fig.1: Structure of Luteolin.

Plants and fruits rich in luteolin such as celery, parsley, broccoli, onion leaves, carrots, peppers, cabbages, apple skins, and chrysanthemum flowers, have been used as Chinese traditional medicine for the treatment of hypertension, inflammatory diseases, and skin cancer [8, 9, 10, 11]. Luteolin is found in plant materials, often in the form of glycosides, which are usually metabolized by the intestinal bacteria, cleaved and glucuronated during uptake in the gut and metabolized in the organism [12]. When it comes to the pharmacological activities of luteolin, it is often associated with its ability to induce apoptosis, which involves redox regulation, DNA damage, protein kinases, which inhibit the proliferation of cancer cells and suppression of metastasis and angiogenesis. Notably, luteolin sensitizes a variety of cancer cells to therapeutically induced cytotoxicity through suppression of cellular pathways and inducing apoptosis pathways. Luteolin is blood-brain barrier

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permeable, rendering it applicable to the therapy of central nerve system diseases, including brain tumor [13]. However, further research is needed in order to fully identify the immunological benefits of this compound.

Biochemical Activities of Luteolin

a) Anti-Carcinogenic Activity: Cancer is the uncontrolled growth of cells, due to mutation. A typical carcinogenic process can be divided into three stages: initiation, promotion, and progression [14]. In the first stage, i.e., the initiation stage, the potential carcinogen (pro-mutagen) is converted to a mutagen by enzymes such as cytochrome P450. The mutagen then reacts with DNA to induce irreversible genetic alteration including mutations, transversions, transitions, and/or small deletions in DNA. In the promotion stage, alterations in genome expression occur to favor cell growth and proliferation. During the progression stage, tumor establishes and becomes irreversible; it is characterized by karyotypic instability and malignant growth in an uncontrolled manner [15]. The transformed cells then acquire a number of characteristic alterations, including the capacity to proliferate in a signal-independent manner, to invade surrounding tissues and metastasize to distant sites. In addition, cancer cells elicit an angiogenic response, evade all mechanisms that limit cell proliferation, such as apoptosis and senescence, and elude immune surveillance [16].

These properties of cancer cells are reflected by alterations in the cellular signaling pathways that control cell proliferation, motility, and survival in normal cells [17]. Luteolin is able to interfere with almost all of the characteristics of cancer cells, mainly through the following mechanisms [18].

a.1) Preventing the activation of Carcinogenic metabolism: From studies done earlier, by various cancer researchers, it was found that luteolin was able to inhibit the metabolism of carcinogens that generates active mutagens in liver microsomes [19, 20]. It was determined recently that luteolin potently inhibits human cytochrome P450 (CYP) 1 family enzymes such as CYP1A1, CYP1A2, and CYP1B1, thereby suppressing the mutagenic activation of carcinogens [21]. By suppressing these enzymes reduces the generation of active mutagens [22].

a.2) Inhibition of Cancer cells growth: Luteolin is able to inhibit the proliferation of cancer cells derived from nearly all types of cancers, as like many other flavonoids, mainly through regulating the cell cycle [23, 24]. In Eukaryotes, cell cycle consists of four distinct phases, G1, S, G2, and M phases, and they are timely regulated by cyclin-dependent kinases (CDKs) and their cyclin subunits at the two checkpoints, G1/S and G2/M [25]. The G1/S checkpoint is regulated by CDK4-cyclin D, CDK6-cyclin D, and CDK2-cyclin E. When associated with cyclin A, CDK2 controls the S-phase, while the G2/M transition is regulated by CDK1 in combination with cyclins A and B, CDK activity is negatively controlled by two groups of CDK inhibitors (CKI), INK4 and CIP/KIP families. The INK4 family members inhibit CDK4 and CDK6; while the CIP/KIP family, consisting of p21cip1/waf1, p27kip1, and p57kip2, inhibits a broad range of CDKs [26].

Various flavonoids have been found to inhibit the proliferation of many cancer cells by arresting cell cycle progression either at the G1/S or G2/M checkpoint [27]. Luteolin is able to arrest the cell cycle during the G1 phase in human gastric and prostate cancer, and in melanoma cells [28, 29, 30]. The G1 cell cycle arrest induced by luteolin is achieved by up-regulation of the CDK inhibitors p27/kip1 and p21/waf1, or direct inhibition on the CDK2 activity [31, 32]. Luteolin arrests mouse cancer cell tsFT210 at the G2/M checkpoint [33]. DNA damage-activated tumor suppressor protein p53 is involved in both the G1/S and G2/M transition regulation [34]. Luteolin can bind and suppress DNA topoisomerases I and II, enzymes essential for repairing damaged DNA, and intercalates directly with the substrate DNA to cause DNA double-strand breaks [35, 36]. This action of luteolin induces cell cycle arrest though p53-mediated expression of p21/waf1 [37].

EGF receptor (EGFR) is a typical receptor protein tyrosine kinase (PTK) that mediates cell growth and proliferation. When activated by its ligands, EGFR is phosphorylated to mediate activation of downstream signaling pathways, including MAPK and PI3K/Akt [38]. Luteolin was found to inhibit the proliferation of pancreatic and prostate cancer and human epidermoid carcinoma cells, which is closely associated with the inhibition of the PTK activity and

autophosphorylation of EGFR, transphosphorylation of EGFR downstream effector protein enolase, and activation of MAPK/ERK [39]. Luteolin is able to inhibit IGF-1-induced activation of IGF-1R and Akt with the suppressed expression of cyclin D1, and it also increased the expression of p21/waf1 and proliferation in prostate cancer cells *in vitro* [40]. In a similar way, luteolin inhibits PDGF-induced proliferation by inhibiting PDGF receptor phosphorylation in vascular smooth muscle cells [41]. As a consequence, luteolin significantly inhibits PDGF-induced ERK, PI3K/Akt and phospholipase C (PLC)- γ 1 activation, and *c-fos* gene expression suggests that the inhibitory effect of luteolin on the PDGF-induced proliferation may be mediated by blocking phosphorylation of the PDGF receptor [42]. As PDGF stimulates cancer cell proliferation, it remains to be determined whether luteolin can block PDGF-induced signaling to suppress cancer cell proliferation [43]. As discussed above, ER induces proliferation in several types of cancer cells [44]. Luteolin suppresses proliferation of prostate and breast cancer cells in both an androgen-dependent and -independent manner, suggesting that luteolin's anti-estrogen activity may at least partly contribute to its anti-proliferation effect [45, 46]. Similar observations were also made in thyroid carcinoma cell lines bearing the ER [47, 48].

In addition to affecting the receptors, luteolin may directly target the downstream pathways that are involved in cell proliferation. For example, protein kinase C, a family of serine-threonine protein kinases that regulates growth factor response and cell proliferation, differentiation and apoptosis [49, 50] can be inhibited in a concentration-dependent manner by luteolin in both cell-free systems and in intact cells [51]. Hence from the reports given above, it was found that the carcinogens activate cell survival pathways such as MAPK during the course of carcinogenesis; these pathways could be additional targets for flavonoids, including luteolin, in anti-carcinogenesis [52, 53].

a.3) Apoptosis of Cancer cells:

Researchers have showed that uncontrolled growth of mutated cells was due to lack of apoptosis is closely associated with tumor formation [54]. The process of apoptosis consists of two pathways, the death receptor

pathway (extrinsic) and the mitochondrial (intrinsic) pathway. The intrinsic pathway involves functional incapacitation of mitochondria by pro-apoptotic Bcl2 family members, including Bax, Bak, and Bik, that cause mitochondria potential loss and release cytochrome c to activate caspase 9, which in turn activates executor caspases (-3, -7) and destroys cellular proteins [55]. The extrinsic pathway is initiated by the binding of TNF family cytokines (TNF α , Fas and TNF-related apoptosis-inducing ligand, TRAIL) to their cognate death receptors, to activate caspase 8, which in turn activates downstream executor caspases [56]. The extrinsic pathway is initiated by the binding of TNF family cytokines (TNF α , Fas and TNF-related apoptosis-inducing ligand, TRAIL) to their cognate death receptors, to activate caspase 8, which in turn activates downstream executor caspases [57, 58]. Despite the complexity underlying luteolin-induced apoptosis, they can still be generalized as breaking the cell survival and death balance by either enhancing apoptosis or decreasing the survival signaling in cancer cells, which is summarized in Fig.2 [59].

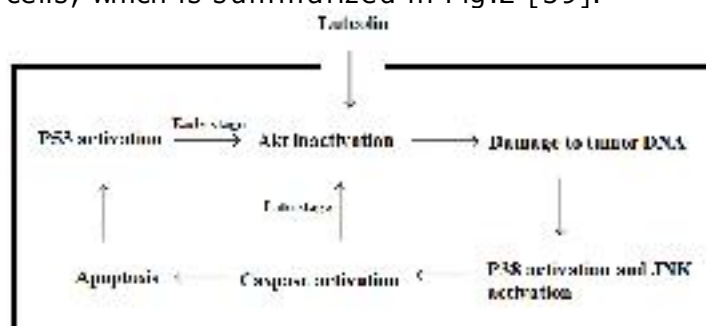


Fig.2: Apoptosis pathways and the points targeted by luteolin.

b) Luteolin as an Anticancer or Chemotherapeutic Agent: As mentioned in previous discussions, luteolin induces apoptosis in various cancer cells [60]. It also has inhibitory effects towards cancer cell growth [61], and suppresses tumor angiogenesis [62]. Supporting the *in vitro* results, *in vivo* experiments in nude mice with xenografted tumors showed that luteolin suppressed growth of tumors formed from human skin carcinoma, hepatoma, and human ovarian cancer cells [63] or mouse Lewis lung carcinoma [64] in a dosage-dependent manner. Luteolin was found to induce marginal cytotoxicity in normal cells [65].

Studies have shown that, the effect of luteolin is enhanced when it is given along with other therapeutic agents, i.e., combined therapy. The drugs tested which were tested along with luteolin include cisplatin [66], TRAIL [66, 66], TNF α [67, 68], and the mTOR inhibitor rapamycin [69]. Luteolin-induced up regulation of the TRAIL receptor DR5 contributes to sensitizing not only TRAIL-induced, but also other chemotherapeutic-induced cytotoxicity [70]. Cancer therapeutics are also found to activate cellular pathways, dampening their cancer cell-killing activities [71, 72]. Yet more research is needed to determine if luteolin or other flavonoids contributes to the anticancer activity of these fruits [73].

In a 20-methylcholanyrene-induced fibrosarcoma model using Swiss albino mice, luteolin administered in diet significantly suppressed tumor incidences, which are associated with reduction in lipid peroxides and cytochrome P450, increased activity of GST, and suppressed DNA synthesis [74]. Luteolin exerts chemopreventive and anticarcinogenic effects, in association with its antiperoxidative and antioxidant effects, against colon cancer [75]. Epidemiological studies suggest that dietary intake of flavonoids is inversely associated with risk of lung, prostate, stomach, and breast cancer in humans [76]. A recent population study on the association between intake of dietary flavonoids and incidence of epithelial ovarian cancer among 66,940 women showed a significant (34%) decrease in cancer incidence for the highest versus lowest luteolin intake [77]. The data suggest that dietary intake of luteolin may reduce ovarian cancer risk, although additional prospective studies are needed [78]. Dietary intake of flavonols and flavones was found to be inversely associated with the risk of lung cancer [79]. However, because of many confounding factors, luteolin's preventive potential for lung cancer still remains unclear [80]. Hence, caution should be exercised when interpreting epidemiological study as well as other results [81].

c) **Other Activities Of Luteolin:**

c.1) **Anti-Inflammatory Activity:**

Inflammation has been one of our body's chief defense mechanisms, to protect us against infections, from pathogens. Yet chronic inflammation may result in harmful diseases such as arthritis, chronic obstructive

pulmonary disease, and cancer [82]. During an infection, there is activation of macrophages due to various components of the system, that vigorously produce inflammatory molecules such as tumor necrosis factor α (TNF α), interleukins (ILs), and free radicals (ROS and reactive nitrogen species, RNS), leading to recruitment of inflammatory cells, such as neutrophils and lymphocytes, to the infection site and clearance of the pathogens [83]. High level production of inflammatory cells, could lead to cancer, hence luteolin is found to exert its anti-inflammatory effect by suppressing the production of these cytokines and their signal transduction pathways [84, 85]. Experiments with animals have showed that luteolin suppresses LPS or bacteria-induced inflammation *in vivo* [86]. LPS-induced-high mortality was effectively alleviated by luteolin, which is associated with reduction of LPS-stimulated TNF α release in serum and intercellular adhesion molecule-1 (ICAM-1) expression in the liver [87]. Luteolin was found to suppress inflammation in lung tissue that was caused by *Chlamydia pneumonia* [88].

Based on the observations that some flavonoids with strong antioxidant activities are completely ineffective in suppressing LPS-stimulated TNF α production, it is assumed that the inhibitory action of flavonoids on proinflammatory cytokine production is not directly associated with their antioxidant properties [89]. However, because luteolin is able to scavenge ROS directly and to suppress the LPS-activated nitric oxide production in activated macrophages, the antioxidant activity of luteolin at least in part contributes to luteolin's anti-inflammatory effect [90, 91]. Because inflammation and its involved signaling pathways are strongly associated with carcinogenesis, the anti-inflammatory role played by luteolin, may contribute to cancer prevention [92, 93].

c.2) Anti-Metastasis Activity: The ability of cancer cells to migrate and invade other, surrounding healthy cells, from their primary location to secondary locations, is called Metastasis. Metastasis, has by far, contributed to over 90% of human cancer mortality [94]. The metastasis cascade is thought to consist of multiple steps: local invasion; intravasation into the systemic circulation; survival during transport, extravasation, and establishment of

micrometastases in distant organs; and colonization of macroscopic metastases [95].

Although direct evidence showing luteolin suppresses cancer metastasis is not seen in literature, available results strongly suggest that luteolin has this function [96]. Luteolin suppresses production and secretion of cytokines such as TNF α and IL-6 that can stimulate cancer cell migration and metastasis [97, 98]. Luteolin blocks the EGFR-signaling pathway and reduces cell invasion and metastasis of tumor cells [99, 100]. Luteolin also directly inhibits the MMP or hyaluronidase enzyme activity to maintain the neovascularization barrier, which may also contribute to suppressing cancer cell metastasis [111, 112]. *In vitro* studies have shown that luteolin potently inhibits migration and invasion of cancer cells through blocking the MAPK/ERKs and PI3K-Akt pathways [113, 114]. Experiments with cancer metastasis animal models are needed to verify luteolin's anti-metastasis activity.

c.3) Anti-Oxidant Activity: Luteolin also possesses excellent anti-oxidant properties. Reactive oxygen species (ROS) refers to a diverse group of reactive, short-lived, oxygen-containing species, such as superoxide ($O_2^{\bullet-}$), hydrogen peroxide (H_2O_2), hydroxyl radical ($\bullet OH$), singlet oxygen (1O_2), and lipid peroxy radical (LOO^{\bullet}). ROS serve as second messengers for cellular signaling [115]. But, the overproduction of ROS was found to cause damage to DNA, lipids, proteins, neuronal disorders, as well as inducing cancer cell production. Luteolin was found to inhibit ROS-induced damage of lipids, DNA, and protein [116, 117]. Multiple mechanisms may contribute to luteolin's antioxidant effect. Firstly, luteolin functions as a ROS scavenger through its own oxidation and it also possesses the structures essential for the flavonoid's antioxidant activity: 3', 4' hydroxylation [118]. Luteolin, due to its aromatic nature, supports unpaired electrons around the M-electron system [119, 120]. Direct evidence showing luteolin as a ROS scavenger was obtained in cell-free systems [121]. Secondly, luteolin suppresses $O_2^{\bullet-}$ formation by inhibiting xanthine oxidase activity, which is a ROS-generating oxidases [122]. But, it is unclear in mammalian cells whether luteolin affects ROS generation in the mitochondria, the main ROS generation site, although it interferes with the mitochondrial electron transportation chain in parasite

(leishmanial) cells [123]. Thirdly, luteolin may exert its antioxidant effect by protecting or enhancing endogenous antioxidants such as glutathione-S-transferase (GST), glutathione reductase (GR), superoxide dismutase (SOD) and catalase (CAT) [124, 125]. Luteolin suppresses lipoxygenase, cyclooxygenase, which are enzymes that catalyze oxidation of the cellular components, and ascorbic acid-stimulated malonaldehyde formation in liver lipids [126]. Finally, luteolin may chelate transition metal ions responsible for the generation of ROS and therefore inhibit lipoxygenase reaction, or suppress nontransition metal-dependent oxidation [127]. It is also noted that, the inhibition of LPS-induced $\bullet OH$ production in macrophages by luteolin may be through scavenging $O_2^{\bullet-}$, inhibiting xanthine oxidase activity, or a combination of both [128].

Discussion

Documented results suggest that luteolin has a variety of beneficial properties, including those as an anti-inflammatory and anticancer agent [129]. The mechanisms underlying these properties have not been fully understood but are partially attributed to luteolin's redox- and regulating properties. It is important and interesting to determine the mechanism for luteolin's selective cytotoxicity in cancerous cells but not normal cells. It is apparent that distinct mechanisms for modulating cellular signaling pathways exist in normal cells and in malignant cancer cells. By understanding the mechanisms, we will undoubtedly facilitate the use of luteolin in cancer prevention and therapy. Although it is relatively safe, luteolin was found to worsen chemically induced colitis in mice [130]. Further studies are needed to address the safety issues of luteolin with doses effective for cancer prevention and therapy in humans [131].

Conclusion

Chronic inflammation is a critical component of tumor development [132]. Multiple lines of evidence suggest that COX-2, a rate-limiting enzyme in the synthesis of prostaglandins, is a key link between inflammation and cancer [133]. Luteolin was found to possess anti-oxidant properties. ROS overproduction was inhibited by luteolin thus, inhibiting the damage of cellular DNA, lipids and proteins and also in the killing of cancer cells. The ROS scavenging activity of luteolin, at least in part, contributes to its nature of

being an anti-inflammatory agent. Through experimental studies, luteolin was found to possess anti-metastasis activity and anti-cancer properties. The Apoptosis pathway and the activation of apoptosis in the cancer cells by luteolin clearly show the anti-carcinogenic effects of luteolin as a therapeutic agent. Luteolin was found to delay the development of tumors, inhibited the multiplication of tumors, decreased tumor volume, and suppressed the expression of inflammatory and proliferative biomarkers.

Luteolin was found to be effective against a number of cancer cells, including skin cancer. The majority of cancers are derived from many mutations [134]. Hence multitargeted kinase inhibitors, like luteolin, are anticipated to have therapeutic advantages and are emerging as new alternatives for cancer treatment [135]. Hence, luteolin was found to possess anti-cancer properties with significant with more animal model studies, to determine the toxicity of luteolin and by effective dosage studies, the compound can be used as a potential cure for cancer.

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References

1. Harborne JB, Williams CA, Advances in flavonoid research since 1992, *Phytochemistry*, 2000, 55, 481–504.
2. Knekt P, Jarvinen R, Seppanen R, Heliovaara M, Teppo L, Pukkala E, Aromaa A, Dietary flavonoids and the risk of lung cancer and other malignant neoplasms, *Am. J. Epidemiol*, 1997, 146, 223–230.
3. Ross JA, Kasum CM, Dietary flavonoids: bioavailability, metabolic effects, and safety, *Annu. Rev. Nutr*, 2002, 22, 19–34.
4. Block G, Patterson B, Subar A, Fruit, vegetables, and cancer prevention: a review of the epidemiological evidence, *Nutr. Cancer*, 1992, 18, 1–29.
5. Birt DF, Hendrich S, Wang W, Dietary agents in cancer prevention: flavonoids and isoflavonoids, *Pharmacol. Ther*, 2001, 90, 157–177.
6. Ross JA, Kasum CM, Dietary flavonoids: bioavailability, metabolic effects, and safety, *Annu. Rev. Nutr*, 2002, 22, 19–34.
7. Chan TS, Galati G, Pannala AS, Rice-Evans C, O'Brien PJ, Simultaneous detection of the antioxidant and pro-oxidant activity of dietary polyphenolics in a peroxidase system, *Free Radic. Res*, 2003, 37, 787–794.
8. Miesan KH, Mohamed S, Flavonoid (myricetin, quercetin, kaempferol, luteolin, and apigenin) content of edible tropical plants, *J. Agric. Food Chem*, 2000, 49, 3106–3112.
9. Gates MA, Tworoger SS, Hecht JL, De Vivo I, Rosner B, Hankinson SE, A prospective study of dietary flavonoid intake and incidence of epithelial ovarian cancer, *Int. J. Cancer*, 2007, 121, 2225–2232.
10. Sun T, Xu Z, Wu CT, Janes M, Prinyawiwatkul W, No HK, Antioxidant activities of different colored sweet bell peppers (*Capsicum annum* L.), *J. Food Sci.* 2007, 72, 98–102.
11. Mencherini T, Picerno P, Scesa C and Aquino R, Triterpene, Antioxidant, and Antimicrobial Compounds from *Melissa officinalis*, *J. Nat. Prod*, 2007, 70, 1889–1894.
12. Hempel J, Pforte H, Raab B, Engst W, Bohm H, Jacobasch G, Flavonols and flavones of parsley cell suspension culture change the antioxidative capacity of plasma in rats, *Nahrung*, 1999, 43, 201–204.
13. Wruck CJ, Claussen M, Fuhrmann G, Romer L, Schulz A, Pufe T, Waetzig V, Peipp M, Herdegen T, Gotz ME, Luteolin protects rat PC12 and C6 cells against MPP+ induced toxicity via an ERK dependent Keap1-Nrf2-ARE pathway, *J. Neural Transm. Suppl*, 2007, 72, 57–67.
14. Pitot HC, Multistage carcinogenesis-genetic and epigenetic mechanisms in relation to cancer prevention, *Cancer Detect Prev*, 1993, 17, 567–573.
15. Hanahan D, Weinberg RA, The hallmarks of cancer, *Cell*, 2000, 100, 57–70.
16. Galati G, Teng S, Moridani MY, Chan TS, O'Brien PJ, Cancer chemoprevention and apoptosis mechanisms induced by dietary polyphenolics, *Drug. Metabol. Drug. Interact*, 2000, 17, 311–349.
17. Buening MK, Chang RL, Huang MT, Fortner JG, Wood AW, Conney AH, Activation and inhibition of benzo (a) pyrene and aflatoxin B1 metabolism in human liver microsomes by naturally occurring flavonoids, *Cancer Res*, 1981, 41, 67–72.

18. Huang MT, Wood AW, Newmark HL, Sayer JM, Yagi H, Jerina DM, Conney AH, Inhibition of the mutagenicity of bay-region diolepoxides of polycyclic aromatic hydrocarbons by phenolic plant flavonoids, *Carcinogenesis*, 1983, 4, 1631 – 1637.
19. Kim HJ, Lee SB, Park SK, Kim HM, Park YI, Dong MS, Effects of hydroxyl group numbers on the B-ring of 5, 7-dihydroxyflavones on the differential inhibition of human CYP 1A and CYP1B1 enzymes, *Arch Pharm Res*, 2005, 28, 1114–1121.
20. Miller KP, Ramos KS, Impact of cellular metabolism on the biological effects of benzo[a]pyrene and related hydrocarbons, *Drug Metab Rev*, 2001, 33, 1–35.
21. Han DH, Denison MS, Tachibana H, Yamada K, Relationship between estrogen receptor-binding and estrogenic activities of environmental estrogens and suppression by flavonoids, *Biosci. Biotechnol. Biochem*, 2002, 66, 1479–1487.
22. Fotsis T, Pepper MS, Aktas E, Breit S, Rasku S, Adlercreutz H, Wahala K, Montesano R, Schweigerer L, Flavonoids, dietary-derived inhibitors of cell proliferation and in vitro angiogenesis, *Cancer Res*, 1997, 57, 2916–2921.
23. Ko WG, Kang TH, Lee SJ, Kim YC, Lee BH, Effects of luteolin on the inhibition of proliferation and induction of apoptosis in human myeloid leukaemia cells, *Phytother Res*, 2002, 16, 295–298.
24. Knowles LM, Zigrossi DA, Tauber RA, Hightower C, Milner JA, Flavonoids suppress androgen-independent human prostate tumor proliferation, *Nutr. Cancer*, 2000, 38, 116–120.
25. Massague J, G1 cell-cycle control and cancer, *Nature*, 2004, 232– 298.
26. Zi X, Feyes DK, Agarwal R, Anticarcinogenic effect of a flavonoid antioxidant, silymarin, in human breast cancer cells MDA-MB 468: induction of G1 arrest through an increase in Cip1/p21 concomitant with a decrease in kinase activity of cyclin-dependent kinases and associated cyclins, *Clin. Cancer Res*, 1998, 4, 1055–1064.
27. Lindenmeyer F, Li H, Menashi S, Soria C, Lu H, Apigenin acts on the tumor cell invasion process and regulates protease production, *Nutr. Cancer*, 2001, 39, 139–147.
28. Matsukawa Y, Marui N, Sakai T, Satomi Y, Yoshida M, Matsumoto K, Nishino H, Aoike A, Genistein arrests cell cycle progression at G2-M, *Cancer Res*, 1993, 53, 1328–1331.
29. Kobayashi T, Nakata T, Kuzumaki T, Effect of flavonoids on cell cycle progression in prostate cancer cells, *Cancer Lett*, 2002, 176, 17–23.
30. Casagrande F, Darbon JM, Effects of structurally related flavonoids on cell cycle progression of human melanoma cells: regulation of cyclin-dependent kinases CDK2 and CDK1, *Biochem. Pharmacol*, 2001, 61, 1205–1215.
31. Lim do Y, Jeong Y, Tyner AL, Park JH, Induction of cell cycle arrest and apoptosis in HT-29 human colon cancer cells by the dietary compound luteolin, *Am J Physiol Gastrointest Liver Physiol*, 2007, 292, 66–75.
32. Li WX, Cui CB, Cai B, Wang HY, Yao XS, Flavonoids from *Vitex trifolia* L, inhibit cell cycle progression at G2/M phase and induce apoptosis in mammalian cancer cells, *J. Asian Nat. Prod. Res*, 2005, 7, 615–626.
33. Caino MC, Oliva JL, Jiang H, Penning TM, Kazanietz MG, Benzo[a]pyrene-7,8-dihydrodiol promotes checkpoint activation and G2/M arrest in human bronchoalveolar carcinoma H358 cells, *Mol. Pharmacol*, 2007, 71, 744–750.
34. Helton ES, Chen X, p53 modulation of the DNA damage response, *J. Cell. Biochem*, 2007, 100, 883–896.
35. Yamashita N, Kawanishi S, Distinct mechanisms of DNA damage in apoptosis induced by quercetin and luteolin, *Free Radic. Res*, 2000, 33, 623–633.
36. Chowdhury AR, Sharma S, Mandal S, Goswami A, Mukhopadhyay S, Majumder HK, Luteolin, an emerging anti-cancer flavonoid, poisons eukaryotic DNA topoisomerase I, *Biochem. J*, 2002, 366, 653–661.
37. Zhang L, Lau YK, Xi L, Hong RL, Kim DS, Chen CF, Hortobagyi GN, Chang C, Hung MC, Tyrosine kinase inhibitors, emodin and its derivative repress HER-2/neu-induced cellular transformation and metastasis-associated properties, *Oncogene*, 1998, 16, 2855–2863.
38. Lee LT, Huang YT, Hwang JJ, Lee PP, Ke FC, Nair MP, Kanadaswam C, Lee MT, Blockade of the epidermal growth factor receptor tyrosine kinase activity by quercetin and luteolin leads to growth inhibition and apoptosis of pancreatic tumor cells, *Anticancer Res*, 2002, 22, 1615–1627.
39. Fang J, Zhou Q, Shi XL, Jiang BH, Luteolin inhibits insulin-like growth factor 1 receptor

- signaling in prostate cancer cells, *Carcinogenesis*, 2007, 28, 713–723.
40. Kim JH, Jin YR, Park BS, Kim TJ, Kim SY, Lim Y, Hong JT, Yoo HS, Yun YP, Luteolin prevents PDGF-BB-induced proliferation of vascular smooth muscle cells by inhibition of PDGF beta-receptor phosphorylation, *Biochem. Pharmacol*, 2005, 69, 1715–1721.
 41. Matei D, Emerson RE, Lai YC, Baldrige LA, Rao J, Yiannoutsos C, Donner DD, Autocrine activation of PDGFR α promotes the progression of ovarian cancer, *Oncogene*, 2006, 25, 2060–2069.
 42. Chiu FL, Lin JK, Downregulation of androgen receptor expression by luteolin causes inhibition of cell proliferation and induction of apoptosis in human prostate cancer cells and xenografts, *Prostate*, 2008, 68, 61–71.
 43. Yin F, Giuliano AE, Van Herle AJ, Growth inhibitory effects of flavonoids in human thyroid cancer cell lines, *Thyroid*, 1999, 9, 369–376.
 44. Lucas M, Sanchez-Margalet V, Protein kinase C involvement in apoptosis, *Gen. Pharmacol*, 1995, 26, 881–887.
 45. Weinstein IB, Kahn SM, O'Driscoll K, Borner C, Bang D, Jiang W, Blackwood A, Nomoto K, The role of protein kinase C in signal transduction, growth control and lipid metabolism, *Adv. Exp. Med. Biol*, 1997, 400A, 313–321.
 46. Ferriola PC, Cody V, Middleton E Jr, Protein kinase C inhibition by plant flavonoids, Kinetic mechanisms and structure-activity relationships, *Biochem. Pharmacol*, 1989, 38, 1617.
 47. Stathopoulos GT, Sherrill TP, Cheng DS, Scoggins RM, Han W, Polosukhin VV, Connelly L, Yull FE, Fingleton B, Blackwell TS, Epithelial NF- κ B activation promotes urethane-induced lung carcinogenesis, *Proc. Natl Acad Sci U.S.A.*, 2007, 104, 18514–18519.
 48. Lee SH, Lee SJ, Kim JH, Park BJ, Chemical carcinogen, N-methyl-N'-nitro-N-nitrosoguanidine, is a specific activator of oncogenic Ras, *Cell Cycle*, 2007, 6, 1257–1264.
 49. Hanahan D, Weinberg RA, The Hallmarks of Cancer, *Cell*, 2000, 100, 57–70.
 50. Wang X, The expanding role of mitochondria in apoptosis, *Genes Dev*, 2001, 15, 2922–2933.
 51. Wajant H, Pfizenmaier K, Scheurich P, Tumor necrosis factor signaling cell death, *Differ*, 2003, 10, 45–65.
 52. Huang YT, Hwang JJ, Lee PP, Ke FC, Huang JH, Huang CJ, Kandaswami C, Middleton E Jr, Lee MT, Effects of luteolin and quercetin, inhibitors of tyrosine kinase, on cell growth and metastasis-associated properties in A431 cells overexpressing epidermal growth factor receptor, *Br J Pharmacol*, 1999, 128, 999–1010.
 53. Cheng A-C, Huang T-C, Lai C-S, Pan M-H, Induction of apoptosis by luteolin through cleavage of Bcl-2 family in human leukemia HL-60 cells, *Eur J Pharmacol*, 2005, 509, 1–10.
 54. Lee HJ, Wang CJ, Kuo HC, Chou FP, Jean LF, Tseng TH, Induction apoptosis of luteolin in human hepatoma HepG2 cells involving mitochondria translocation of Bax/Bak and activation of JNK, *Toxicol Appl Pharmacol*, 2005, 203, 124–131.
 55. Lin Y, Shi RX, Wang X, Shen HM, Luteolin, a flavonoid with potential for cancer prevention and therapy, *Curr Cancer Drug Targets*, 2008, 8, 634–646.
 56. Horinaka M, Yoshida T, Shiraishi T, Nakata S, Wakada M, Nakanishi R, Nishino H, Matsui H, Sakai T, Luteolin induces apoptosis via death receptor 5 upregulation in human malignant tumor cells, *Oncogene*, 2005, 24, 7180–7189.
 57. Selvendiran K, Koga H, Ueno T, Yoshida T, Maeyama M, Torimura T, Yano H, Kojiro M, Sata M, Luteolin promotes degradation in signal transducer and activator of transcription 3 in human hepatoma cells: an implication for the antitumor potential of flavonoids, *Cancer Res*, 2006, 66, 4826–4834.
 58. Plaumann B, Fritsche M, Rimpler H, Brandner G, Hess RD, Flavonoids activate wild-type p53, *Oncogene*, 1996, 13, 1605–1614.
 59. Shi R, Huang Q, Zhu X, Ong YB, Zhao B, Lu J, Ong CN, Shen HM, Luteolin sensitizes the anticancer effect of cisplatin via c-Jun NH₂-terminal kinase-mediated p53 phosphorylation and stabilization, *Mol Cancer Ther*, 2007, 6, 1338–1347.
 60. Ju W, Wang X, Shi H, Chen W, Belinsky SA, Lin Y, A critical role of luteolin-induced reactive oxygen species in blockage of tumor necrosis factor-activated nuclear factor- κ B pathway and sensitization of apoptosis in lung cancer cells, *Mol Pharmacol*, 2007, 71, 1381–1388.
 61. Shi RX, Ong CN, Shen HM, Luteolin sensitizes tumor necrosis factor- α -induced apoptosis in human tumor cells, *Oncogene*, 2004, 23, 7712–7721.

62. Yu C, Minemoto Y, Zhang J, Liu J, Tang F, Bui TN, Xiang J, Lin A, JNK suppresses apoptosis via phosphorylation of the proapoptotic Bcl-2 family protein BAD, *Mol Cell*, 2004, 13, 329–340.
63. Greten FR, Eckmann L, Greten TF, Park JM, Li ZW, Egan LJ, Kagnoff MF, Karin M, IKK β links inflammation and tumorigenesis in a mouse model of colitis-associated cancer, *Cell*, 2004, 118, 285–296.
64. Malhi H, Gores GJ, TRAIL resistance results in cancer progression: a TRAIL to perdition? *Oncogene*, 2006, 25, 7333–7335.
65. Shi R-X, Ong C-N, Shen H-M, Protein Kinase C Inhibition and X-Linked Inhibitor of Apoptosis Protein Degradation Contribute to the Sensitization Effect of Luteolin on Tumor Necrosis Factor-Related Apoptosis-Inducing Ligand-Induced Apoptosis in Cancer Cells, *Cancer Res*, 2005, 65, 7815–7823.
66. Chang J, Hsu Y, Kuo P, Kuo Y, Chiang L, Lin C, Increase of Bax/Bcl-XL ratio and arrest of cell cycle by luteolin in immortalized human hepatoma cell line, *Life Sci*, 2005, 76, 1883–1893.
67. Brusselmans K, Vrolix R, Verhoeven G, Swinnen JV, Induction of cancer cell apoptosis by flavonoids is associated with their ability to inhibit fatty acid synthase activity, *J Biol Chem*, 2005, 280, 5636–5645.
68. Lupu R, Menendez JA, Pharmacological inhibitors of Fatty Acid Synthase (FASN)--catalyzed endogenous fatty acid biogenesis: a new family of anticancer agents? *Curr Pharm Biotechnol*, 2006, 7, 483–493.
69. Monasterio A, Urdaci MC, Pinchuk IV, Lopez-Moratalla N, Martinez-Irujo JJ, Flavonoids induce apoptosis in human leukemia U937 cells through caspase- and caspase-calpain-dependent pathways, *Nutr Cancer*, 2004, 50, 90–100.
70. Chiang CT, Way TD, Lin JK, Sensitizing HER2-overexpressing cancer cells to luteolin-induced apoptosis through suppressing p21(WAF1/CIP1) expression with rapamycin, *Mol Cancer Ther*, 2007, 6, 2127–2138.
71. Kandaswami C, Lee LT, Lee PP, Hwang JJ, Ke FC, Huang YT, Lee MT, The antitumor activities of flavonoids, *In Vivo*, 2005, 19, 895–909.
72. Bagli E, Stefanidou M, Morbidelli L, Ziche M, Psillas K, Murphy C, Fotsis T, Luteolin inhibits vascular endothelial growth factor-induced angiogenesis; inhibition of endothelial cell survival and proliferation by targeting phosphatidylinositol 3'-kinase activity, *Cancer Res*, 2004, 64, 7936–7946.
73. Kim JH, Lee EO, Lee HJ, Ku JS, Lee MH, Yang DC, Kim SH, Caspase activation and extracellular signal-regulated kinase/Akt inhibition were involved in luteolin-induced apoptosis in Lewis lung carcinoma cells, *Ann N Y Acad Sci*, 2007, 1095, 598–611.
74. Samy RP, Gopalakrishnakone P, Ignacimuthu S, Anti-tumor promoting potential of luteolin against 7,12-dimethylbenz(a)anthracene-induced mammary tumors in rats, *Chem Biol Interact*, 2006, 164, 1–14.
75. Chen D, Chen MS, Cui QC, Yang H, Dou QP, Structure-proteasome-inhibitory activity relationships of dietary flavonoids in human cancer cells, *Front Biosci*, 2007, 12, 1935–1945.
76. Friesen C, Fulda S, Debatin KM, Cytotoxic drugs and the CD95 pathway, *Leukemia*, 1999, 13, 1854–1858.
77. Belyanskaya LL, Hopkins-Donaldson S, Kurtz S, Simoes-Wüst AP, Yousefi S, Simon HU, Stahel R, Zangemeister-Wittke U, Cisplatin activates Akt in small cell lung cancer cells and attenuates apoptosis by survivin upregulation, *Int J Cancer*, 2005, 117, 755–763.
78. Lin Y, Devin A, Cook A, Keane MM, Kelliher M, Lipkowitz S, Liu ZG, The death domain kinase RIP is essential for TRAIL (Apo2L)-induced activation of IkappaB kinase and c-Jun N-terminal kinase, *Mol Cell Biol*, 2000, 20, 6638–6645.
79. Huang C, Huang Y, Li J, Hu W, Aziz R, Tang MS, Sun N, Cassady J, Stoner GD, Inhibition of benzo(a)pyrene diol-epoxide-induced transactivation of activated protein 1 and nuclear factor kappaB by black raspberry extracts, *Cancer Res*, 2002, 62, 6857–6863.
80. Lu H, Li J, Zhang D, Stoner GD, Huang C, Molecular mechanisms involved in chemoprevention of black raspberry extracts: from transcription factors to their target genes, *Nutr Cancer*, 2006, 54, 69–78.
81. Li J, Zhang D, Stoner GD, Huang C, Differential effects of black raspberry and strawberry extracts on BaPDE-induced activation of transcription factors and their target genes, *Mol Carcinog*, 2008, 47, 286–294.
82. Raina K, Singh RP, Agarwal R, Agarwal C, Oral grape seed extract inhibits prostate tumor growth and progression in TRAMP mice, *Cancer Res*, 2007, 67, 5976–5982.

83. Veeriah S, Kautenburger T, Habermann N, Sauer J, Dietrich H, Will F, Pool-Zobel BL, Apple flavonoids inhibit growth of HT29 human colon cancer cells and modulate expression of genes involved in the biotransformation of xenobiotics, *Mol Carcinog*, 2006, 45, 164–174.
84. Elangovan V, Sekar N, Govindasamy S, Chemopreventive potential of dietary bioflavonoids against 20-methylcholanthrene-induced tumorigenesis, *Cancer Lett*, 1994, 87, 107–113.
85. Osakabe N, Yasuda A, Natsume M, Yoshikawa T, Rosmarinic acid inhibits epidermal inflammatory responses: anticarcinogenic effect of *Perilla frutescens* extract in the murine two-stage skin model, *Carcinogenesis*, 2004, 25, 549–557.
86. Ueda H, Yamazaki C, Yamazaki M, Inhibitory effect of *Perilla* leaf extract and luteolin on mouse skin tumor promotion, *Biol Pharm Bull*, 2003, 26, 560–563.
87. Manju V, Nalini N, Protective role of luteolin in 1,2-dimethylhydrazine induced experimental colon carcinogenesis, *Cell Biochem Funct*, 2007, 25, 189–194.
88. Knekt P, Jarvinen R, Seppanen R, Heliovaara M, Teppo L, Pukkala E, Aromaa A, Dietary flavonoids and the risk of lung cancer and other malignant neoplasms, *Am J Epidemiol*, 1997, 146, 223–230.
89. Wright ME, Mayne ST, Stolzenberg-Solomon RZ, Li Z, Pietinen P, Taylor PR, Virtamo J, Albanes D, Development of a comprehensive dietary antioxidant index and application to lung cancer risk in a cohort of male smokers, *Am J Epidemiol*, 2004, 160, 68–76.
90. Hirvonen T, Virtamo J, Korhonen P, Albanes D, Pietinen P, Flavonol and flavone intake and the risk of cancer in male smokers (Finland), *Cancer Causes Control*, 2001, 12, 789–796.
91. Garcia-Closas R, Agudo A, Gonzalez CA, Riboli E, Intake of specific carotenoids and flavonoids and the risk of lung cancer in women in Barcelona, Spain, *Nutr Cancer*, 1998, 32, 154–158.
92. Brody JS, Spira A, State of the art, Chronic obstructive pulmonary disease, inflammation, and lung cancer, *Proc Am Thorac Soc*, 2006, 3, 535–537.
93. Perwez Hussain S, Harris CC, Inflammation and cancer: an ancient link with novel potentials, *Int J Cancer*, 2007, 121, 2373–2380.
94. Karin M, Lawrence T, Nizet V, Innate immunity gone awry: linking microbial infections to chronic inflammation and cancer, *Cell*, 2006, 124, 823–835.
95. Xagorari A, Papapetropoulos A, Mauromatis A, Economou M, Fotsis T, Roussos C, Luteolin inhibits an endotoxin-stimulated phosphorylation cascade and proinflammatory cytokine production in macrophages, *J Pharmacol Exp Ther*, 2001, 296, 181–187.
96. Chen C-C, Chow M-P, Huang W-C, Lin Y-C, Chang Y-J, Flavonoids Inhibit Tumor Necrosis Factor- α -Induced Up-Regulation of Intercellular Adhesion Molecule-1 (ICAM-1) in Respiratory Epithelial Cells through Activator Protein-1 and Nuclear Factor- κ B: structure-Activity relationships, *Mol Pharmacol*, 2004, 66, 683–693.
97. Kumazawa Y, Kawaguchi K, Takimoto H, Immunomodulating effects of flavonoids on acute and chronic inflammatory responses caused by tumor necrosis factor α , *Curr Pharm Des*, 2006, 12, 4271–4279.
98. Kotanidou A, Xagorari A, Bagli E, Kitsanta P, Fotsis T, Papapetropoulos A, Roussos C, Luteolin Reduces Lipopolysaccharide-induced Lethal Toxicity and Expression of Proinflammatory Molecules in Mice, *Am J Respir Crit Care Med*, 2002, 165, 818–823.
99. Tormakangas L, Vuorela P, Saario E, Leinonen M, Saikku P, Vuorela H, In vivo treatment of acute *Chlamydia pneumoniae* infection with the flavonoids quercetin and luteolin and an alkyl gallate, octyl gallate, in a mouse model, *Biochem Pharmacol*, 2005, 70, 1222–1230.
100. Devasagayam TP, Subramanian M, Singh BB, Ramanathan R, Das NP, Protection of plasmid pBR322 DNA by flavonoids against single-stranded breaks induced by singlet molecular oxygen, *J Photochem Photobiol B*, 1995, 30, 97–103.
101. Kim HK, Cheon BS, Kim YH, Kim SY, Kim HP, Effects of naturally occurring flavonoids on nitric oxide production in the macrophage cell line RAW 264,7 and their structure-activity relationships, *Biochem Pharmacol*, 1999, 58, 759–765.
102. Hu C, Kitts DD, Luteolin and luteolin-7-O-glucoside from dandelion flower suppress iNOS and COX-2 in RAW264, 7 cells, *Mol Cell Biochem*, 2004, 265, 107–113.
103. Chen CY, Peng WH, Tsai KD, Hsu SL, Luteolin suppresses inflammation-associated gene expression by blocking NF- κ B and

- AP-1 activation pathway in mouse alveolar macrophages, *Life Sci*, 2007, 81, 1602-1614.
104. Beuvink I, Boulay A, Fumagalli S, Zilbermann F, Ruetz S, O'Reilly T, Natt F, Hall J, Lane HA, Thomas G, The mTOR Inhibitor RAD001 Sensitizes Tumor Cells to DNA-Damaged Induced Apoptosis through Inhibition of p21 Translation, *Cell*, 2005, 120, 747-759.
105. Szlosarek P, Charles KA, Balkwill FR, Tumour necrosis factor-alpha as a tumour promoter, *Eur J Cancer*, 2006, 42, 745-750.
106. Yang J, Mani SA, Weinberg RA, Exploring a new twist on tumor metastasis, *Cancer Res*, 2006, 66, 4549-4552.
107. Chen CY, Peng WH, Tsai KD, Hsu SL, Luteolin suppresses inflammation-associated gene expression by blocking NF-kappaB and AP-1 activation pathway in mouse alveolar macrophages, *Life Sci*, 2007, 81, 1602-1614.
108. Beuvink I, Boulay A, Fumagalli S, Zilbermann F, Ruetz S, O'Reilly T, Natt F, Hall J, Lane HA, Thomas G, The mTOR Inhibitor RAD001 Sensitizes Tumor Cells to DNA-Damaged Induced Apoptosis through Inhibition of p21 Translation, *Cell*, 2005, 120, 747-759.
109. Szlosarek P, Charles KA, Balkwill FR, Tumour necrosis factor-alpha as a tumour promoter, *Eur J Cancer*, 2006, 42, 745-750.
110. Lee L-T, Huang Y-T, Hwang J-J, Lee AYL, Ke F-C, Huang C-J, Kandaswami C, Lee P-PH, Lee M-T, Transinactivation of the epidermal growth factor receptor tyrosine kinase and focal adhesion kinase phosphorylation by dietary flavonoids: effect on invasive potential of human carcinoma cells, *Biochem Pharmacol*, 2004, 67, 2103-2114.
111. Kuppusamy UR, Khoo HE, Das NP, Structure-activity studies of flavonoids as inhibitors of hyaluronidase, *Biochem Pharmacol*, 1990, 40, 397-401.
112. Ende C, Gebhardt R, Inhibition of matrix metalloproteinase-2 and -9 activities by selected flavonoids, *Planta Med*, 2004, 70, 1006-1008.
113. Lansky EP, Harrison G, Froom P, Jiang WG, Pomegranate (*Punica granatum*) pure chemicals show possible synergistic inhibition of human PC-3 prostate cancer cell invasion across Matrigel, *Invest New Drugs*, 2005, 23, 121-122.
114. Lee WJ, Wu LF, Chen WK, Wang CJ, Tseng TH, Inhibitory effect of luteolin on hepatocyte growth factor/scatter factor-induced HepG2 cell invasion involving both MAPK/ERKs and PI3K-Akt pathways, *Chem Biol Interact*, 2006, 160, 123-133.
115. Robak J, Shridi F, Wolbis M, Krolukowska M, Screening of the influence of flavonoids on lipoxygenase and cyclooxygenase activity, as well as on nonenzymic lipid oxidation, *Pol J Pharmacol Pharm*, 1988, 40, 451-458.
116. Brown JE, Rice-Evans CA, Luteolin-rich artichoke extract protects low density lipoprotein from oxidation in vitro, *Free Radic Res*, 1998, 29, 247-255.
117. Lien EJ, Ren S, Bui HH, Wang R, Quantitative structure-activity relationship analysis of phenolic antioxidants, *Free Radic Biol Med*, 1999, 26, 285-294.
118. Shimoi K, Masuda S, Furugori M, Esaki S, Kinae N, Radioprotective effect of antioxidative flavonoids in gamma-ray irradiated mice, *Carcinogenesis*, 1994, 15, 2669-2672.
119. Nagao A, Seki M, Kobayashi H, Inhibition of xanthine oxidase by flavonoids, *Biosci Biotechnol Biochem*, 1999, 63, 1787-1790.
120. Sen N, Das BB, Ganguly A, Banerjee B, Sen T, Majumder HK, Leishmania donovani: intracellular ATP level regulates apoptosis-like death in luteolin induced dyskinetoplastid cells, *Exp Parasitol*, 2006, 114, 204-214.
121. Leung HW, Kuo CL, Yang WH, Lin CH, Lee HZ, Antioxidant enzymes activity involvement in luteolin-induced human lung squamous carcinoma CH27 cell apoptosis, *Eur J Pharmacol*, 2006, 534, 12-18.
122. Manju V, Nalini N, Chemopreventive potential of luteolin during colon carcinogenesis induced by 1,2-dimethylhydrazine, *Ital J Biochem*, 2005, 54, 268-275.
123. Nimnual AS, Taylor LJ, Bar-Sagi D, Redox-dependent downregulation of Rho by Rac, *Nat Cell Biol*, 2003, 5, 236-241.
124. Harris GK, Qian Y, Leonard SS, Sbarra DC, Shi X, Luteolin and chrysin differentially inhibit cyclooxygenase-2 expression and scavenge reactive oxygen species but similarly inhibit prostaglandin-E2 formation in RAW 264, 7 cells, *J Nutr*, 2006, 136, 1517-1521.
125. Xagorari A, Roussos C, Papapetropoulos A, Inhibition of LPS-stimulated pathways in macrophages by the flavonoid luteolin, *Br J Pharmacol*, 2002, 136, 1058-1064.
126. Kim JS, Jobin C, The flavonoid luteolin prevents lipopolysaccharide-induced NF- κ B signalling and gene expression by blocking I κ B kinase activity in intestinal epithelial cells and

- bone-marrow derived dendritic cells, *Immunology*, 2005, 115, 375–387.
127. Karrasch T, Kim JS, Jang BI, Jobin C, The flavonoid luteolin worsens chemical-induced colitis in NF-kappaB (EGFP) transgenic mice through blockade of NF-kappaB-dependent protective molecules, *PLoS ONE*, 2007, 2, e596.
 128. Coussens LM, Werb Z, Inflammation and cancer, *Nature*, 2002, 420, 860–867.
 129. DuBois RN, Abramson SB, Crofford L, Cyclooxygenase in biology and disease, *FASEB J*, 1998, 12, 1063–1073.
 130. Prescott SM, Fitzpatrick FA, Cyclooxygenase-2 and carcinogenesis, *Biochim Biophys Acta Rev*, 2000, 1470.
 131. Balkwill F, Coussens LM, Cancer: an inflammatory link, *Nature*, 2004, 431, 405–406.
 132. Kim Y, Fischer SM, Transcriptional regulation of cyclooxygenase-2 in mouse skin carcinoma cells: regulatory role of CCAAT/enhancerbinding proteins in the differential expression of cyclooxygenase-2 in normal and neoplastic tissues, *J Biol Chem*, 1998, 273, 27686–27694.
 133. Tang Q, Gonzales M, Inoue H, Bowden GT, Roles of akt and glycogen synthase kinase 3 β in the ultraviolet B induction of cyclooxygenase-2 transcription in human keratinocytes, *Cancer Res*, 2001, 61, 4329–4332.
 134. Bode AM, Dong Z, Signal transduction pathways: targets for chemoprevention of skin cancer, *Lancet Oncol*, 2000, 1, 181–188.
 135. Dong Z, Birrer MJ, Watts RG, Matrisian LM, Colburn NH, Blocking of tumor promoter-induced AP-1 activity inhibits induced transformation in JB6 mouse epidermal cells, *Proc Natl Acad Sci USA*, 1994, 91, 609–613.

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